

INTELLECTUAL DISABILITY CLASSIFICATION, CAUSES, EPIGENETIC MECHANISMS AND TREATMENT

ANJUM R¹, REHMAN AU², MAQSOOD H², ILYAS U², NIAZ M³, ROHAIL⁴, MOHSIN S^{1*}, JURRAT H¹, ANJUM S⁵, MUNAWAR I⁴, HAMID M^{6*}, ZAFAR MZ^{2*}

 ¹ Department of Zoology, Government College University, Lahore, Pakistan, ²Faisalabad Medical University, Allied Hospital Faisalabad, Pakistan
³Department of Zoology, Wildlife and Fisheries, University of Agriculture, Faisalabad, Pakistan ⁴Department of Zoology, Riphah International University, Faisalabad Campus, Pakistan ⁵Shaukat Khanum Memorial Cancer Hospital & Research Centre, Pakistan ⁶Registrar, Family Physician, King Saud University Medical City, Riyadh, Saudi Arabia *Correspondence author email address: <u>sehrishmohsin@gmail.com</u>; <u>mashod@live.com</u>; <u>zafarzubairrana@gmail.com</u>

(Received, 11th November 2022, Revised 16th April 2023, Published 21st April 2023)

Abstract: Intellectual disability (ID) is a condition characterized by a defective adaptive and cognitive attitudes that can occur with various mental disorders, such as attention-deficit/hyperactivity and an autism spectrum disorder. It may also be associated with malformation syndromes affecting other organs. Genetic studies have linked several chromatin-modifying enzymes and epigenetic regulators to ID disorders (IDDs). This review explores how dysfunction in histone modifiers, chromatin remodelers, and methyl-DNA binding proteins can cause neurodevelopmental deformities and alter brain plasticity. The use of mouse models generated through human genetics has allowed researchers to uncover the molecular basis of ID and explore potential therapeutic strategies. Understanding the chromatin regulators associated with IDDs has broader implications for treating other IDDs, as they target common downstream genes and cellular functions. Investigating these disorders can also shed light on the function of chromatin regulators in brain growth, plasticity, and gene regulation, leading to new insights into fundamental questions in neurobiology.

Keywords: Intellectual disability, Epigenetic, Malformation syndrome

Introduction

Intellectual disability (ID) can result in incomplete or arrested mental development. It is characterized by the deterioration of cognitive, language, motor, and socialization functions at each stage of development, leading to an overall low level of intelligence. Individuals with ID struggle to adapt to their environment due to the pervasive impact of their cognitive impairments. Generally, a person with an intelligence quotient (IQ) of 70 or below has two or more behavioural deficits. The IQ is the standard test developed to assess the disability in humans (Selmen et al., 2005). In ID, the patient finds difficulty in daily activities such as communication, personal care, sociability, self-governance, health and safety, and academic skills (Matson et al., 2005). It is mostly diagnosed before the age of 18 years when a person is unable to perform properly or is diagnosed with adaptive functioning (Gorgoni et al., 2020). The ID Occurrence rate is 1 to 3% of the total population and independent of social stratum (Maulik et al., 2011).

Instead of the universal data, it is shown that ID is more common in the lower socioeconomic status and developing areas, where its severity occurs from mild to worse depending on the treatment (Durkin, 2002; Emerson, 2007). ID is likely to occur because of Ecological factors (Luckasson, & Schalock, 2013; Emerson, 2002), but genetic factors contribute equally.

In the general population, about 30% more males are affected by ID thano females. But it is believed that as IQ decreases, the prevalence of ID alsdecreasesed (American Psychiatric Association, 2002). Previous research showed that severe ID is more prevalent in females than males (Bradley et al., 2002).

Classification of ID

ID is classified into four general categories that is mild, moderate, severe and profound (Matson *et al.*, 2005). But some epidemiological studies classified ID into two categories: mild ID with IQ 50-70 and severe ID with IQ<50 (Ropers and Hamel, 2005). The

[Citation: Anjum, R., Rehman, A.U., Maqsood, H., Ilyas, U., Niaz, M., Rohail., Mohsin, S., Jurrat, H., Anjum, S., Munawar, I., Hamid, M., Zafar, M.Z. (2023). Intellectual disability classification, causes, epigenetic mechanisms and treatment. *Biol. Clin. Sci. Res. J.*, **2023**: 245. doi: <u>https://doi.org/10.54112/bcsrj.v2023i1.245</u>]

1



occurrence of ID differs depending upon the study design, and the age of the subject caused the variabilities (Leonard and Wen, 2002), as shown in Table 1.

ID's severity can be divided into syndromic intellectual disability (SID) and non-syndromic intellectual disability (NSID). In the SID, the patient usually has one more clinical feature: co-morbidities

besides ID. While the NSID, the patient has ID as the only clinical feature. But to create the boundary between SID and NSID is difficult. Diagnosing one or more clinical features (neurological and psychiatric abnormalities) in these patients is very difficult to rule out. Also, the ID syndromes are so subtle that they are very difficult to find as they may be linked to genetic defects (Ropers, 2006).

Table 1| Classification of intellectual disability from mild to profound.

Levels of ID	IQ Range	Academic Skills	Communication and Language	Basic Skills	Support and Care
Mild (85%)	50-69	6 th -7 th grade	Able to do simple multiplication, division, writing letters and form lists.	Able to do self-care and home activities. Use public transport.	Support is short term. Can have independent living.
Moderate (10%)	35-49	2 nd -3 rd grade	Sight word reading, copy addresses from card, item numbers.	Able to do some self-care and home activities. Use public transport with supervision.	Require consistent support but can survive for little.
Severe (5%)	20-34	Primary level	Able to identify own things.	Due to motor impairments, required daily support. Able to do some daily activities.	Require consistent support and care dependent.
Profound (>3%)	Below 20	None	Did not do anything due to motor impairments.	Many motor and sensory impairments. Require daily support.	Require full time support with high supervision.

Causes of ID

The environmental, genetic factors or both contribute to ID. There are about 60% unknown cases of ID which remain unidentified (Rauch *et al.*, 2006). The environmental causes of ID include exposure to toxins, teratogens, viruses or radiation, which can damage (severe head trauma or injury) brain cells and cause a lack of oxygen in the brain tissues, which eventually die and causes more damage to brain. These factors mostly occurred due to the lack of awareness in underdeveloped or developing countries.

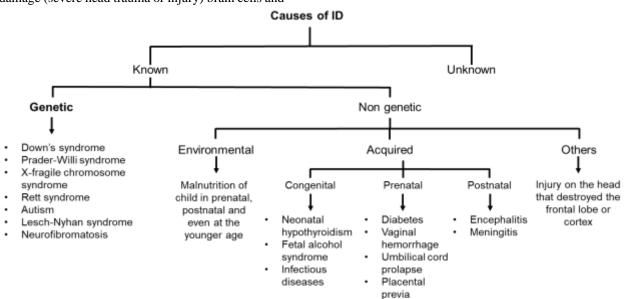


Figure 1] Causes of intellectual disability (ID) with examples. ID is caused by genetic and non-genetic factors. In the genetic factors, X-fragile chromosome syndrome, autism and down's syndrome are most common. On the other hand, the non-genetic factors are the environmental such as malnutrition or acquired in any stage of pregnancy or the head trauma even in the adult life causes the ID. Many other factors contribute to ID which are still unknown.

Genetic factors

Genetic defects cause the majority of cases of intellectual disability (ID). Specifically, about 25-50% cases of ID are attributed to genetic defects, and this proportion tends to increase with the intensity of the ID (McLaren and Bryson, 1987). The most common cause of ID is chromosomal abnormalities, and numerous abnormalities are found (Rauch et al., 2006). Autosomal trisomies and X-chromosome aneuploidies are often associated with intellectual disability (ID) in humans due to their link to genetic instability. Down syndrome, caused by trisomy 21, is a frequent form of ID. Numerous genetic components have been recognized that participate in the development of ID (Zahir and Friedman, 2007). The less common forms of chromosomal abnormalities include X-fragile chromosome syndrome (Miclea et al., 2015), Prader-Willi syndrome (Jauregi et al., 2007), Rett syndrome (Weaving et al., 2005), neurofibromatosis (Mouridsen and Sorensen, 1995). tuberous sclerosis (Curatolo et al., 2015). Lesch-Nyhan syndrome (Nyhan et al., 1989) and adrenoleukodystrophy (Feanny et al., 1987) rarely occur (Figure 1). In the last 15 years, many forms of NSID were identified due to environmental and genetic factors. Autism or neurodevelopment disorder is likely to be occurred by genetic basics, or environmental factors may contribute equally. The NSID is multifactorial, with one or more genes involved in the disease. It can either be autosomal or X-linked (Chelly et al., 2006).

Acquired factors

Acquired factors may be congenital or developmental. The congenital factors include the neonatal metabolic hypothyroidism, toxins such as lead poisoning (Tellez-Rojo *et al.*, 2006), fetal alcohol syndrome (Kodituwakku, 2007) and prenatal exposure to substances (Morrow *et al.*, 2006), and the infectious diseases such as rubella (Desmond *et al.*, 1969), syphilis (Gilad *et al.*, 2007), toxoplasmosis (Tomita *et al.*, 2021) and simple herpes genital type II (Seppanen *et al.*, 2006).

During pregnancy, in the prenatal period, complexities such as uncontrolled diabetes (Leonard *et al.*, 2006), intrauterine malnutrition (Calis *et al.*, 2007), vaginal hemorrhages (Schellae *et al.*, 1994), placenta previa (Naeye, 1978) and umbilical cord prolapse (Niswander *et al.*, 1975). Also, in this period, prolonged suffering from anoxia and asphyxia related suffocation also retard the growth of brain tissues (Slitonen *et al.*, 2003). In postnatal period, infectious diseases like encephalitis and meningitis affects the fetus (Noyola *et al.*, 2001) shown in figure 2.

Environmental and socioeconomic factors

Many epidemiological studies showed that intellectual disability mostly related to family status. There are many notable links between ID and poverty. Firstly, poverty exposes the person to many environmental and psychotic stimulus (Leonard *et al.*, 2005). Second, if a person in the poor family has ID, it more causes a burden on the other family members. These factors are mostly affected in the developing countries (Emerson and Hatton, 2007). These interactions result in the child's malnutrition in prenatal, postnatal and even at the younger age.

Epigenetic mechanisms in ID

The ID is mostly affected by epigenesis, which is the not the change in the DNA sequence but only regulated the chromatin state in the DNA (Waddington, 1956). These mechanisms are interrelated, but chromatin compaction is the only thing common in them. Impaired transcription results in the loose of the facilitated gene. The molecular mechanisms still need to be explained. Epigenetic mechanisms participate in coordination among genes and the environment, particularly in learning and memory processes. These mechanisms contribute to brain plasticity, which involves modifying neuronal structures in response to external inputs. Recent studies have demonstrated that neural impulses initiate the production of new proteins in dendrites, which in turn affect the function of postsynaptic neurons (Cajigas et al., 2010). This review highlights the epigenetic dysregulation in ID.

1. Chemical modification of DNA

An enzyme, DNA methyltransferases (DNMTs) catalyze the DNA modifications. DNMTs usually transfer the methyl group from the S-adenyl methionine (SAM) to the cytosine residues to form the 5-methyl cytosine (5mC). Cytosine methylation usually occurs at the CpG site and then 5mC demethylated the thymine (Bird, 1980). These sites are found on the gene promotors regions, which are highly conserved andhave high density of CpG sites (>50%). In general, these sites usually interfere with the transcription binding factors and repress the methyl binding domains of proteins (Bird et al., 1980; Nan et al., 1993).

In mammals, three different types of DNMTs exist: DNMT1 is the maintenance DMNT, as it binds to hemi-methylated sites and prevents demethylation during the DNA synthesis. DMNT3a and DMNT3b are the de novo DMNTs (Okanno et al., 1999). As a fact, DMNTs are abundantly expressed in the brain not only in the neurodevelopmental stage but also in the postmitotic neurons, which explains their role beyond the DNA methylation (Feng et al., 2005). The DNA methylation is a static epigenetic process that can only be affected by the demethylation during the cell division. So, the DNA methylation should be regulated. Ten-eleven translocations (TET) enzymes usually oxidise the 5mC. In the next step, it is deaminated by the AID/Apobec enzymes, which oxidise the TET enzymes. In the end, the oxidized product is repaired by BER (Ito et al., 2011).

Many studies show that DNA methylation is related to intellectual by involving the genes. When considering the contextual fear conditioning in the rodent model to study memory; during memory formation, DNMTs are elevated in hippocampus, resulting in increased DNA methylation at the promoter of the memory suppressor gene PP1, and decreased methylation at the promoter of the synaptic plasticity gene RELN throughout memory consolidation (Miller and Swett, 2007). This also happens with the BDNF gene, in which DNA methylation occurs in the learning task, which results in increased BDNF exon I and IV in the fear memory consolidation (Lubin et al., 2008). The DNA methylation changes in the brain are transientand revert in 24 hours. This suggests that DNA methylation have significant involvement in the formation and consolidation of memories in the hippocampus (Figure 2)

Additionally, the brain exhibits elevated amounts of two additional categories of methylation: non-CpG methylation (mCH, where H represents adenine A, cvtosine C, or thymine T) and hydroxymethylation (5hmC), both of which are involved in neuronal modifications (Varley et al., 2013; Song et al., 2011). mCH is not present in the fetal cortex but accumulates during postnatal development, leading to DNA methylation and gene repression in neurodevelopment. From this point, DNMT3a plays an important role in the neurons during development. Higher level of mCH in the glial cells causes the suppression of neuron-specific gene and methylated at the level of CH in the glial cells (Lister et al., 2013). Overexpression of TET1 causes disrupts the formation of contextual fear memories. (Kass et al., 2013), while the TET1 downregulation leads to deficiencies in synaptic plasticity and memory excitation (Rudenko et al., 2013).

Numerous researchers reported the function of DNA methylation in the hippocampus during memory formation and consolidation but have not explored the long-term storage of memories. It has been established that a burst of waves, known as sharp waves, promotes plasticity, facilitating the transfer of memories from the hippocampus to the neocortex (Wiltgen et al., 2004). These waves result in the epigenetic storing of learning in the cortical cells by DNA methylation and resistance to erasing the DNMT1 self-perpetuating mechanism, which usually methylated the hemi-methylated and unmethylated stands of DNA (Heyward and Sweatt, 2015). The CaN (calcineurin) gene is involved in the maintenance of memories, as delayed to persistent DNA methylation occurs in cortical neurons for 1 to over 30 days, respectively, causing contextual fear memory to transition from transient to remote (Miller et al., 2010).

2. Histone modification

The primary constituents of chromatin are histones, which can be categorized into four major types: H4, H2A, H3, and H2B. These histones correlated tightly with DNA to form a nucleosome. H1 controls the folding of the nucleosome, while posttranslational modifications (PTMs) govern chromatin compaction by modifying the protruding end of the histone tail (Bannister and Kouzarides et al., 2011). Several PMTs act on the tail of histone such as acetylation, methylation, phosphorylation, SUMOylation and ADP-ribosylation (Shin et al., 2015).

Histone acetylation usually causes a positive effect on chromatin folding by neutralizing acetyl group from the residues of lysine (K) and arginine (R), thus reducing the electrostatic interactions in the DNA nucleosome. These epigenetic writers are known as histone acetyltransferases (HATs), and the erasers known as histone deacetylases (HDACs) (Lopez and Barco, 2014). This histone acetvlation causes memory regulation by the ERK/MAP pathway regulation in the cortex by lysine acetylation (Levenson et al., 2004). Many types of research revealed the HDACs inhibitor (HDADi) improves the cognitive impairment and increase memory and learning (Lopez and Barco, 2014). These modification causes the transcription and prepares the cells to initiate the gene regulation on signals (Lopez-Atlaya et al., 2013). CREB is a specific transcriptional factor that coactivates the CBP through HAT domain, increases the acetylation process at gene level, and helps in memory integration (Korzus et al., 2004)(Figure 2).

Many HDAC isoforms regulate the adult form, specifically at the histone acetylation level. For example, HDAC5 causes the hypersensitive response to chronic drug abuse (Tsankova et al., 2006); HDAC2 causes the deregulation in memory formation and synaptic plasticity (Guan et al., 2009), and HDAC3 inhibits long-term memory formation (McQuown et al., 2011). SIRT1 impairs the hippocampal formation of memory by decreasing the dendritic branching and spines, which is the special structure in learning (Michan et al., 2010). HDCA1 requires the deacetylation of H3K9 for the fear extinction learning (Bahari-Javan et al., 2012) and HDAC4 requires memory formation and synaptic plasticity (Kim et al., 2012).

The process of histone phosphorylation is closely linked to histone acetylation. Specifically, the phosphorylation of histone H3 at the serine (S) 10 site (H3S10P) occurs alongside the acetylation of histone H3K9. This biochemical event plays a critical role in forming spatial memory and activating many genes (such as c-Fos, Erg-1, and Arc) by the ERK/MAPK pathway (Carter et al., 2015).

The histone acetylation results in transcriptional regulation, while the methylation of histone effects mostly depends upon the docking protein complexes.

For instance, the H3K4 methylation and monomethylation of H3K9 cause transcriptional activation, while H3K9me2 and H3K9ma3 cause transcriptional knockdown. Histone methylation occurs either at lysine (K) or arginine (R) is mostly carried out by the SET of proteins domain known as histone methyl transferase (HMTs). HMTs are regulated by the histone demethylases (HDMs), such as LSD1 for H3K4me and H3K4me2 and JMJD1a for H3K9me and H3K9me2 (Lubin et al., 2011).

Recent study in which mice deficit with Mll, a H3K4 methyltransferase, shows defects in fear memory formation (Gupta et al., 2010). GLP/G9a, an H3K9me2 methyltransferase, plays an important role in cognition and switching chromatin signals (Benevento et al., 2015), acting during development and modulating the gene expression by recruiting the enzymes. This complex helps in memory consolidation from hippocampus to cortex (Gupta et al., 2012). GLP/G9a is also important in developing the behaviour because its deficiency cause the defects in learning, memory and motivation (Schaefer et al., 2009).

3. Chromatin remodeling

Nucleosome remodeling complexes (NRCs) use ATP-dependent mechanisms to change the position of nucleosomes by enhancing nucleosome sliding, expulsion, and exchange of histone variants. Studies have shown that the neuron-specific Brg1/hBrm associated factor (nBAF) complex, which belongs to the SWI/SNF family, is involved in activating gene expression during both development and cognition. Upregulation of BAF45b and BAF45c subunits and BAF53a and BAF53b is essential in postmitotic neurons (Figure 2) and control the BRG1's ATPase activity (Olave et al., 2012). The mice deficit with BAF53b showed large impairments in long-term memory consolidation (Vogel-Ciernia et al., 2013).

4. Noncoding RNAs (ncRNAs)

ncRNAs are part of the transcript that are not translated to proteins. It has two main categories: small RNAs and long noncoding RNAs (lncRNAs). The first comprehends the micro RNA (miRNA), which inhibits the gene expression at the target and interacts with PIWI interacting RNAs (piRNAs) and suppresses RNA mediated DNA methylation. The lncRNAs role is not well known but it regulates the gene expression by guiding and scaffolding RNAs and targeting the genomic locations (Cao et al., 2006). TUNA, RMST, and DALI regulate neuronal differentiation, which guide transcription factors, chromatin remodelling, and DNMTs to crucial locations (Chalei et al., 2014; Kerioglu et al., 2013). In many cases, methyl-binding proteins bring in HDACs, which work together through cytosine methylation and histone deacetylation to silence gene transcription (Vaissiere et al., 2008). By comparing these mechanisms, it can be said that gene expressions controlled brain activity by controlling the DNA methylation, histone acetylation and chromatin remodelling. (Figure 2)

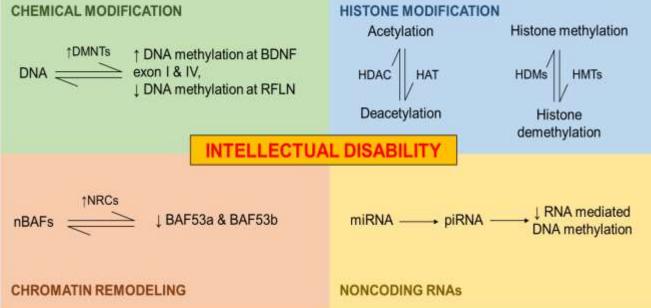


Figure 2| Epigenetic mechanisms in the intellectual disability; (1) chemical modification results in the methylation of DNA, (2) histone modifications causes the deacetylation and demethylation od the histone molecules, (3) in chromatin remodeling the BAF decreased results in ID and (4) noncoding RNAs decreased the RNA based DNA methylation. DMNT (DNA methyltransferases); HDAC (Histone deacetylase); HAT (Histone acetyltransferase); BAF(Brg1/hBrm associated factor); NRC (nucleosome remodeling complexes); miRNA (micro RNA); piRNA (PIWI RNA).

Treatment of ID

ID is now considered a neurodevelopmental disorder rather than just focusing on the intellectual level. A patient may adapt to existing environment if the positive reinforcements are given to them continuously. As ID cannot be cured completely, behavior is normalized according to society.

1. Environment enrichment

In this review, previously, it is pointed out that environment has a strong influence on epigenetic modifications. The microenvironment affects or interrupts the genome as any stage of development in any area of brain. The monozygotic pairs could show this effect, which are genetically similar but phenotypically changed if they drifted apart (Castillo-Fernandez et al., 2014). Environment enrichment (EE) is mostly used in rodent models to enhance learning and memory. For example, keeping the lab mice in EE with large cages and many toys to develop their sensory, cognitive and motor skills. The results showed that mice living in EE had improved learning and memory (Grill et al., 2009), and delayed deficits neurological disorders mouse model in (Nithianantharajah and Hannan, 2006). EE did not interfere with the genome but only improve the dendritic branching and spines as shown in Down's syndrome model (Dierssen et al., 2003). In ID, the EE may cause some modification in the epigenetic mechanisms, as the EE only lasts for 3-4 weeks while behavioural changes are lifetime (Shin et al., 2013). Four weeks training in the EE conditions to rescue the contextual fear conditioning and water maze assays, there is an increased histone acetylation residues (Fischer et al., 2007).

Understanding the complete mechanism to cure ID, a new class of therapeutics was developed known as enviro mimetics. Environimetics mimics the EE beneficial effects on learning (Nithianantharajah and Hannan, 2006). Many research using EE paves the way for the nonpharmacological treatment of ID.

2. Epigenetic drugs in ID

To understand ID and its treatment, a thorough gene profile was developed to know the key genes causing the impairments (Schaefer et al., 2015). A most promising way to last the transcriptional changes is by using cancer research (Dawson and Kouzarides, 2012). Epigenetic changes are reversible so alleviating some gene expressions will help control the ID. FDA already approves some drugs, two DNMT inhibitors (5-azacytidine and decitabine) and two HDAC inhibitors (Nebbioso et al., 2012). Also, the valproic acid, already utilized against the epilepsy and bipolar defects. It revealed the HDAC inhibition, anticholinergic action, and the first epidrug used in neurological disorders (Papi et al., 2010). But in using these types of drugs caused the genome-wide and nonchromatin effect as they may interfere with the nonhistone proteins. So, much research are conducted to find the specific drugs that only target the particular gene (de Groote *et al.*, 2012).

Conflict of interest

The authors declared absence of conflict of interest. **References**

- Ambalavanan, N., Carlo, W. A., Shankaran, S., Bann, C. M., Emrich, S. L., Higgins, R. D., ... & Donovan, E. F. (2006). Predicting outcomes of neonates diagnosed with hypoxemic-ischemic encephalopathy. *Pediatrics*, **118**(5), 2084-2093.
- American Psychiatric Association. 2000. Diagnostic and statistical manual of mental disorders, Text Revision. 4th ed. Washington: American Psychiatric Association.
- Bahari-Javan, S., Maddalena, A., Kerimoglu, C., Wittnam, J., Held, T., Bähr, M., ... & Sananbenesi, F. (2012). HDAC1 regulates fear extinction in mice. *Journal of Neuroscience*, **32**(15), 5062-5073.
- Bannister, A. J., & Kouzarides, T. (2011). Regulation of chromatin by histone modifications. *Cell research*, **21**(3), 381.
- Benevento, M., van de Molengraft, M., van Westen, R., van Bokhoven, H., & Kasri, N. N. (2015). The role of chromatin repressive marks in cognition and disease: A focus on the repressive complex GLP/G9a. *Neurobiology* of learning and memory, **124**, 88-96.
- Bird, A. P. (1980). DNA methylation and the frequency of CpG in animal DNA. *Nucleic acids research*, **8**(7), 1499-1504.
- Bird, A., Taggart, M., Frommer, M., Miller, O. J., & Macleod, D. (1985). A fraction of the mouse genome that is derived from islands of nonmethylated, CpG-rich DNA. *Cell*, 40(1), 91-99.
- Bradley, E. A., Thompson, A., & Bryson, S. E. (2002). Mental retardation in teenagers: prevalence data from the Niagara region, Ontario. *The Canadian Journal of Psychiatry*, **47**(7), 652-659.
- Cajigas, I. J., Will, T., & Schuman, E. M. (2010). Protein homeostasis and synaptic plasticity. *The EMBO journal*, **29**(16), 2746-2752.
- Calis, E. A., Olieman, J. F., Rieken, R., & Penning, C. (2007). Impact of malnutrition on gastrointestinal disorders and gross motor abilities in children with cerebral palsy. *Brain and Development*, **29**(8), 534.
- Cao, X., Yeo, G., Muotri, A. R., Kuwabara, T., & Gage, F. H. (2006). Noncoding RNAs in the mammalian central nervous system. *Annu. Rev. Neurosci.*, 29, 77-103.
- Carter, S. D., Mifsud, K. R., & Reul, J. M. (2015). Distinct epigenetic and gene expression changes in rat hippocampal neurons after

- Castillo-Fernandez, J. E., Spector, T. D., & Bell, J. T. (2014). Epigenetics of discordant monozygotic twins: implications for disease. *Genome medicine*, **6**(7), 60.
- Chalei, V., Sansom, S. N., Kong, L., Lee, S., Montiel, J. F., Vance, K. W., & Ponting, C. P. (2014). The long non-coding RNA Dali is an epigenetic regulator of neural differentiation. *Elife*, *3*, e04530.
- Chelly, J., Khelfaoui, M., Francis, F., Chérif, B., & Bienvenu, T. (2006). Genetics and pathophysiology of mental retardation. *European Journal of Human Genetics*, **14**(6), 701.
- Dawson, M. A., & Kouzarides, T. (2012). Cancer epigenetics: from mechanism to therapy. *cell*, **150**(1), 12-27.
- de Groote, M. L., Verschure, P. J., & Rots, M. G. (2012). Epigenetic editing: targeted rewriting of epigenetic marks to modulate expression of selected target genes. *Nucleic acids research*, **40**(21), 10596-10613.
- Desmond, M. M., Montgomery, J. R., Melnick, J. L., Cochran, G. G., & Verniaud, W. (1969). Congenital rubella encephalitis: effects on growth and early development. *American Journal of Diseases of Children*, **118**(1), 30-31.
- Dierssen, M., Benavides-Piccione, R., Martínez-Cué, C., Estivill, X., Flórez, J., Elston, G. N., & DeFelipe, J. (2003). Alterations of neocortical pyramidal cell phenotype in the Ts65Dn mouse model of Down syndrome: effects of environmental enrichment. *Cerebral cortex*, **13**(7), 758-764.
- Drews, C. D., Yeargin-Allsopp, M., Decoufle, P., & Murphy, C. C. (1995). Variation in the influence of selected sociodemographic risk factors for mental retardation. *American journal of public health*, **85**(3), 329-334.
- Durkin, M. (2002). The epidemiology of developmental disabilities in low-income countries. *Mental retardation and developmental disabilities research reviews*, 8(3), 206-211.
- Emerson, E. (2007). Poverty and people with intellectual disabilities. *Mental retardation and developmental disabilities research reviews*, **13**(2), 107-113.
- Emerson, E., & Hatton, C. (2007). Contribution of socioeconomic position to health inequalities of British children and adolescents with intellectual disabilities. *American Journal on Mental Retardation*, **112**(2), 140-150.
- Feanny, S. J., Chuang, S. H., Becker, L. E., & Clarke, J. T. R. (1987). Intracerebral paraventricular

hyperdensities: a new CT sign in Krabbe globoid cell leukodystrophy. *Journal of inherited metabolic disease*, **10**(1), 24-27.

- Feng, J., Chang, H., Li, E., & Fan, G. (2005). Dynamic expression of de novo DNA methyltransferases Dnmt3a and Dnmt3b in the central nervous system. *Journal of neuroscience research*, **79**(6), 734-746.
- Fischer, A., Sananbenesi, F., Wang, X., Dobbin, M., & Tsai, L. H. (2007). Recovery of learning and memory is associated with chromatin remodelling. *Nature*, **447**(7141), 178.
- Gilad, R., Lampl, Y., Blumstein, G., & Dan, M. (2007). Neurosyphilis: the reemergence of an historical disease. *IMAJ-RAMAT GAN-*, **9**(2), 117.
- Grilli, M., Zappettini, S., Zanardi, A., Lagomarsino, F., Pittaluga, A., Zoli, M., & Marchi, M. (2009). Exposure to an enriched environment selectively increases the functional response of the presynaptic NMDA receptors which modulate noradrenaline release in mouse hippocampus. *Journal of Neurochemistry*
- Guan, J. S., Haggarty, S. J., Giacometti, E., Dannenberg, J. H., Joseph, N., Gao, J., ... & Bradner, J. E. (2009). HDAC2 negatively regulates memory formation and synaptic plasticity. *Nature*, **459**(7243), 55.
- Guo, J. U., Su, Y., Zhong, C., Ming, G. L., & Song, H. (2011). Hydroxylation of 5-methylcytosine by TET1 promotes active DNA demethylation in the adult brain. *Cell*, **145**(3), 423-434.
- Gupta, S., Kim, S. Y., Artis, S., Molfese, D. L., Schumacher, A., Sweatt, J. D., ... & Lubin, F. D. (2010). Histone methylation regulates memory formation. *Journal of Neuroscience*, **30**(10), 3589-3599.
- Gupta-Agarwal, S., Franklin, A. V., DeRamus, T., Wheelock, M., Davis, R. L., McMahon, L. L., & Lubin, F. D. (2012). G9a/GLP histone lysine dimethyltransferase complex activity in the hippocampus and the entorhinal cortex is required for gene activation and silencing during memory consolidation. *Journal of Neuroscience*, **32**(16), 5440-5453.
- Heyward, F. D., & Sweatt, J. D. (2015). DNA methylation in memory formation: emerging insights. *The Neuroscientist*, 21(5), 475-489.
- Ito, S., Shen, L., Dai, Q., Wu, S. C., Collins, L. B., Swenberg, J. A., ... & Zhang, Y. (2011). Tet proteins can convert 5-methylcytosine to 5formylcytosine and 5carboxylcytosine. *Science*, 333(6047), 1300-1303.
- Jauregi, J., Arias, C., Vegas, O., Alen, F., Martinez, S., Copet, P., & Thuilleaux, D. (2007). A neuropsychological assessment of frontal cognitive functions in Prader–Willi

syndrome. *Journal of Intellectual Disability Research*, **51**(5), 350-365.

- Kaas, G. A., Zhong, C., Eason, D. E., Ross, D. L., Vachhani, R. V., Ming, G. L., ... & Sweatt, J. D. (2013). TET1 controls CNS 5methylcytosine hydroxylation, active DNA demethylation, gene transcription, and memory formation. *Neuron*, **79**(6), 1086-1093.
- Katusic, S. K., Colligan, R. C., Beard, C. M., O'Fallon, W. M., Bergstralh, E. J., Jacobsen, S. J., & Kurland, L. T. (1996). Mental retardation in a birth cohort, 1976-1980, Rochester, Minnesota. American journal of mental retardation: AJMR, 100(4), 335-344.
- Kerimoglu, C., Agis-Balboa, R. C., Kranz, A., Stilling, R., Bahari-Javan, S., Benito-Garagorri, E., ... & Fischer, A. (2013). Histone-methyltransferase MLL2 (KMT2B) is required for memory formation in mice. *Journal of Neuroscience*, 33(8), 3452-3464.
- Kim, M. S., Akhtar, M. W., Adachi, M., Mahgoub, M., Bassel-Duby, R., Kavalali, E. T., ... & Monteggia, L. M. (2012). An essential role for histone deacetylase 4 in synaptic plasticity and memory formation. *Journal of Neuroscience*, **32**(32), 10879-10886.
- Kodituwakku, P. W. (2007). Defining the behavioral phenotype in children with fetal alcohol spectrum disorders: a review. *Neuroscience & Biobehavioral Reviews*, **31**(2), 192-201.
- Korzus, E., Rosenfeld, M. G., & Mayford, M. (2004). CBP histone acetyltransferase activity is a critical component of memory consolidation. *Neuron*, **42**(6), 961-972.
- Leonard, H., & Wen, X. (2002). The epidemiology of mental retardation: challenges and opportunities in the new millennium. *Mental* retardation and developmental disabilities research reviews, **8**(3), 117-134.
- Leonard, H., de Klerk, N., Bourke, J., & Bower, C. (2006). Maternal health in pregnancy and intellectual disability in the offspring: a population-based study. *Annals of epidemiology*, **16**(6), 448-454.
- Leonard, H., Petterson, B., De Klerk, N., Zubrick, S. R., Glasson, E., Sanders, R., & Bower, C. (2005). Association of sociodemographic characteristics of children with intellectual disability in Western Australia. *Social science* & *medicine*, **60**(7), 1499-1513.
- Levenson, J. M., O'Riordan, K. J., Brown, K. D., Trinh, M. A., Molfese, D. L., & Sweatt, J. D. (2004). Regulation of histone acetylation during memory formation in the hippocampus. *Journal of Biological Chemistry*, **279**(39), 40545-40559.

- Lin, N., Chang, K. Y., Li, Z., Gates, K., Rana, Z. A., Dang, J., ... & Head, S. R. (2014). An evolutionarily conserved long noncoding RNA TUNA controls pluripotency and neural lineage commitment. *Molecular cell*, 53(6), 1005-1019.
- Lister, R., Mukamel, E. A., Nery, J. R., Urich, M., Puddifoot, C. A., Johnson, N. D., ... & Yu, M. (2013). Global epigenomic reconfiguration during mammalian brain development. *Science*, **341**(6146), 1237905.
- Lopez-Atalaya, J. P., & Barco, A. (2014). Can changes in histone acetylation contribute to memory formation? *Trends in Genetics*, **30**(12), 529-539.
- Lopez-Atalaya, J. P., Ito, S., Valor, L. M., Benito, E., & Barco, A. (2013). Genomic targets, and histone acetylation and gene expression profiling of neural HDAC inhibition. *Nucleic acids research*, **41**(17), 8072-8084.
- Lubin, F. D., Gupta, S., Parrish, R. R., Grissom, N. M., & Davis, R. L. (2011). Epigenetic mechanisms: critical contributors to long-term memory formation. *The Neuroscientist*, **17**(6), 616-632.
- Lubin, F. D., Roth, T. L., & Sweatt, J. D. (2008). Epigenetic regulation of BDNF gene transcription in the consolidation of fear memory. *Journal of Neuroscience*, 28(42), 10576-10586.
- Matson, J. L., Dixon, D. R., Matson, M. L., & Logan, J. R. (2005). Classifying mental retardation and specific strength and deficit areas in severe and profoundly mentally retarded persons with the MESSIER. *Research in developmental disabilities*, 26(1), 41-45.
- McLaren, J., & Bryson, S. E. (1987). Review of recent epidemiological studies of mental retardation: prevalence, associated disorders, and etiology. *American Journal on Mental Retardation*.
- McLaren, J., & Bryson, S. E. (1987). Review of recent epidemiological studies of mental retardation: prevalence, associated disorders, and etiology. *American Journal on Mental Retardation*.
- McQuown, S. C., Barrett, R. M., Matheos, D. P., Post, R. J., Rogge, G. A., Alenghat, T., ... & Wood, M. A. (2011). HDAC3 is a critical negative regulator of long-term memory formation. *Journal of Neuroscience*, **31**(2), 764-774.
- Michán, S., Li, Y., Chou, M. M. H., Parrella, E., Ge, H., Long, J. M., ... & Mervis, R. F. (2010). SIRT1 is essential for normal cognitive function and synaptic plasticity. *Journal of Neuroscience*, 30(29), 9695-9707.

- Miller, C. A., & Sweatt, J. D. (2007). Covalent modification of DNA regulates memory formation. *Neuron*, **53**(6), 857-869.
- Miller, C. A., Gavin, C. F., White, J. A., Parrish, R. R., Honasoge, A., Yancey, C. R., ... & Sweatt, J. D. (2010). Cortical DNA methylation maintains remote memory. *Nature neuroscience*, **13**(6), 664.
- Morrow, C. E., Culbertson, J. L., Accornero, V. H., Xue, L., Anthony, J. C., & Bandstra, E. S. (2006). Learning disabilities and intellectual functioning in school-aged children with prenatal cocaine exposure. *Developmental neuropsychology*, **30**(3), 905-931.
- Mouridsen, S. E., & Sørensen, S. A. (1995). Psychological aspects of von Recklinghausen neurofibromatosis (NF1). *Journal of medical* genetics, **32**(12), 921-924.
- Naeye, R. L. (1978). Placenta previa. Predisposing factors and effects on the fetus and surviving infants. *Obstetrics and gynecology*, **52**(5), 521-525.
- Nan, X., Meehan, R. R., & Bird, A. (1993). Dissection of the methyl-CpG binding domain from the chromosomal protein MeCP2. *Nucleic acids research*, **21**(21), 4886-4892.
- Nebbioso, A., Carafa, V., Benedetti, R., & Altucci, L. (2012). Trials with 'epigenetic'drugs: an update. *Molecular oncology*, **6**(6), 657-682.
- Niswander, K. R., Gordon, M., & Drage, J. S. (1975). The effect of intrauterine hypoxia on the child surviving to 4 years. *American journal of* obstetrics and gynecology, **121**(7), 892-899.
- Nithianantharajah, J., & Hannan, A. J. (2006). Enriched environments, experience-dependent plasticity and disorders of the nervous system. *Nature Reviews Neuroscience*, **7**(9), 697.
- Noyola, D.E., Demmler, G.J., Nelson, C.T., Griesser, C., Williamson, W.D., Atkins, J.T., Rozelle, J., Turcich, M., Llorente, A.M., Sellers-Vinson, S. and Reynolds, A., (2001). Early predictors of neurodevelopmental outcome in symptomatic congenital cytomegalovirus infection. *The Journal of pediatrics*, **138**(3), 325-331.
- Nyhan, W. L., Wulfeck, B. B., Tallal, P., & Marsden, D. L. (1989). Metabolic correlates of learning disability. *Birth defects original article series*, **25**(6), 153-169.
- Okano, M., Bell, D. W., Haber, D. A., & Li, E. (1999). DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development. *Cell*, **99**(3), 247-257.
- Olave, I., Wang, W., Xue, Y., Kuo, A., & Crabtree, G. R. (2002). Identification of a polymorphic, neuron-specific chromatin remodeling

complex. *Genes* & *development*, **16**(19), 2509-2517.

- Papi, A., Ferreri, A. M., Rocchi, P., Guerra, F., & Orlandi, M. (2010). Epigenetic modifiers as anticancer drugs: effectiveness of valproic acid in neural crest-derived tumor cells. *Anticancer research*, **30**(2), 535-540.
- Rauch, A., Hoyer, J., Guth, S., Zweier, C., Kraus, C., Becker, C., Zenker, M., Hüffmeier, U., Thiel, C., Rüschendorf, F. and Nürnberg, P., 2006. Diagnostic yield of various genetic approaches in patients with unexplained developmental delay or mental retardation. *American journal* of medical genetics Part A, **140**(19), pp.2063-2074.
- Raznahan, A., Higgins, N. P., Griffiths, P. D., Humphrey, A., Yates, J. R., & Bolton, P. F. (2007). Biological markers of intellectual disability in tuberous sclerosis. *Psychological medicine*, **37**(9), 1293-1304.
- Reiss, A. L., & Hall, S. S. (2007). Fragile X syndrome: assessment and treatment implications. *Child and adolescent psychiatric clinics of North America*, **16**(3), 663-675.
- Roeleveld, N., Zielhuis, G. A. and Gabreels, F. (1997). The prevalence of mental retardation: a critical review of recent literature. *Developmental Medicine & Child Neurology*, **39**(2), 125-132.
- Ropers, H. H. (2006). X-linked mental retardation: many genes for a complex disorder. *Current* opinion in genetics & development, **16**(3), 260-269.
- Ropers, H. H., & Hamel, B. C. (2005). X-linked mental retardation. *Nature reviews genetics*, **6**(1), 46.
- Rudenko, A., Dawlaty, M. M., Seo, J., Cheng, A. W., Meng, J., Le, T., ... & Tsai, L. H. (2013). Tet1 is critical for neuronal activity-regulated gene expression and memory extinction. *Neuron*, **79**(6), 1109-1122.
- Schaefer, A., Sampath, S. C., Intrator, A., Min, A., Gertler, T. S., Surmeier, D. J., ... & Greengard, P. (2009). Control of cognition and adaptive behavior by the GLP/G9a epigenetic suppressor complex. *Neuron*, **64**(5), 678-691.
- Schaefer, T. L., Davenport, M. H., & Erickson, C. A. (2015). Emerging pharmacologic treatment options for fragile X syndrome. *The application of clinical genetics*, **8**, 75.
- Scheller, J. M., & Nelson, K. B. (1994). Does cesarean delivery prevent cerebral palsy or other neurologic problems of childhood? *Obstetrics and gynecology*, **83**(4), 624-630.
- Selman, V., Selman, R. C., Selman, J., & Selman, E. (2005). Spiritual-intelligence/-

- Seppänen, M., Meri, S., Notkola, I. L., Seppälä, I. J., Hiltunen-Back, E., Sarvas, H., ... & Lokki, M. L. (2006). Subtly impaired humoral immunity predisposes to frequently recurring genital herpes simplex virus type 2 infection and herpetic neuralgia. *The Journal of infectious diseases*, **194**(5), 571-578.
- Sharif, M., Ziaei, H., Daryani, A., & Ajami, A. (2007). Seroepidemiological study of toxoplasmosis in intellectual disability children in rehabilitation centers of northern Iran. *Research in developmental disabilities*, 28(3), 219-224.
- Shin, J., Ming, G. L., & Song, H. (2015). Seeking a roadmap toward neuroepigenetics. *Neuron*, **86**(1), 12-15.
- Shin, S. S., Bales, J. W., Yan, H. Q., Kline, A. E., Wagner, A. K., Lyons-Weiler, J., & Dixon, C. E. (2013). The effect of environmental enrichment on substantia nigra gene expression after traumatic brain injury in rats. *Journal of neurotrauma*, **30**(4), 259-270.
- Siitonen, S. L., Kauppinen, T., Leino, T. K., Vanninen, E., Kuronen, P., & Länsimies, E. (2003). Cerebral blood flow during acceleration in flight measured with SPECT. Aviation, space, and environmental medicine, 74(3), 201-206.
- Song, C. X., Szulwach, K. E., Fu, Y., Dai, Q., Yi, C., Li, X., ... & Wang, J. (2011). Selective chemical labeling reveals the genome-wide distribution of 5hydroxymethylcytosine. *Nature biotechnology*, **29**(1), 68.
- Téllez-Rojo, M. M., Bellinger, D. C., Arroyo-Quiroz, C., Lamadrid-Figueroa, H., Mercado-Garcia, A., Schnaas-Arrieta, L., ... & Hu, H. (2006). Longitudinal associations between blood lead concentrations lower than 10 μg/dL and neurobehavioral development in environmentally exposed children in Mexico City. *Pediatrics*, **118**(2), e323-e330.
- Tsankova, N. M., Berton, O., Renthal, W., Kumar, A., Neve, R. L., & Nestler, E. J. (2006). Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. *Nature neuroscience*, **9**(4), 519.
- Vaissière, T., Sawan, C., & Herceg, Z. (2008). Epigenetic interplay between histone modifications and DNA methylation in gene silencing. *Mutation Research/Reviews in Mutation Research*, 659(1-2), 40-48.
- Varley, K. E., Gertz, J., Bowling, K. M., Parker, S. L., Reddy, T. E., Pauli-Behn, F., ... & Absher, D. M. (2013). Dynamic DNA methylation across

diverse human cell lines and tissues. *Genome* research, **23**(3), 555-567.

- Vogel-Ciernia, A., Matheos, D. P., Barrett, R. M., Kramár, E. A., Azzawi, S., Chen, Y., ... & Jia, Y. (2013). The neuron-specific chromatin regulatory subunit BAF53b is necessary for synaptic plasticity and memory. *Nature neuroscience*, **16**(5), 552.
- Waddington, C. H. (1956). Genetic assimilation of the bithorax phenotype. *Evolution*, **10**(1), 1-13.
- Weaving, L. S., Ellaway, C. J., Gecz, J., & Christodoulou, J. (2005). Rett syndrome: clinical review and genetic update. *Journal of medical genetics*, 42(1), 1-7.
- Wiltgen, B. J., Brown, R. A., Talton, L. E., & Silva, A. J. (2004). New circuits for old memories: the role of the neocortex in consolidation. *Neuron*, 44(1), 101-108.
- Zahir, F., & Friedman, J. M. (2007). The impact of array genomic hybridization on mental retardation research: a review of current technologies and their clinical utility. *Clinical genetics*, **72**(4), 271-287.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licen ses/by/4.0/. © The Author(s) 2023