

COMPUTATIONAL ANALYSIS OF CHILLI INFECTING PEPPER LEAF CURL LAHOREVIRUS AND PEPPER LEAF CURL BANGLADESH VIRUS

ATIF M<sup>1</sup>, AHMAD F<sup>2</sup>, MANZOOR MT<sup>1\*</sup>, GILANI K<sup>3</sup>, ALI Q<sup>4</sup>, SARWAR M<sup>5</sup>, ANJUM S<sup>6</sup>, ALAM MW<sup>7</sup>, HUSSAIN A<sup>8</sup>, RAFAQAT N<sup>1</sup>

<sup>1</sup>Department of Plant Pathology, Faculty of Agricultural Sciences, University of the Punjab, Lahore, Pakistan

<sup>2</sup>School of Food Sciences and Technology, Minhaj University Lahore, Pakistan

<sup>3</sup>Institute of Molecular Biology and Biotechnology, University of Lahore, Lahore, Pakistan

<sup>4</sup>Department of Plant Breeding and Genetics, Faculty of Agricultural Sciences, University of the Punjab, Lahore, Pakistan

<sup>5</sup>Department of Horticulture, Faculty of Agricultural Sciences, University of the Punjab, Lahore, Pakistan

<sup>6</sup>Department of Botany, University of the Punjab, Lahore, Pakistan

<sup>7</sup>Department of Plant Pathology, University of Okara, Okara, Pakistan

<sup>8</sup>Department of Agronomy, Faculty of Agricultural Sciences, University of the Punjab, Lahore, Pakistan

\*Corresponding author's email: [tariq.iags@pu.edu.pk](mailto:tariq.iags@pu.edu.pk)

(Received, 15<sup>th</sup> May 2022, Revised 8<sup>th</sup> October 2022, Published 9<sup>th</sup> October 2022)

**Abstract:** Since a few decades ago, the pandemic of the Chilli Leaf Curl Disease Complexes (ChiLCD) has caused enormous alarm. The investigations of various genomes isolated from various regions of the subcontinent in the current study revealed a vast expansion of ChiLCD. According to evolutionary research, genomes from the pepper leaf curl Lahore virus (PepLCLV) (JN880419 and JX524173) and the pepper leaf curl Bangladesh virus (PepLCBV) (DQ116881 and KY420149) are ancestors while others are descendants. The accessions JN880419, JX524173, DQ116881, and KY420149's sequences underwent evolution, giving rise to several strains and variations. According to the findings, the Indian subcontinent's chilli crop may be most severely affected by the extremely aggressive ChiLCV.

**Keywords:** ChiLCD, Diversity, Geminiviruses, Mutation, Phylogenetic analysis

### Introduction

Begomoviruses are generally significant and most devastating infections, spread by whiteflies (*Bemisia tabaci*). Around thirty years, begomoviruses infections have been arose significant pathogen in different crops like brinjal, papaya and American cotton etc. It's possible that the genomes of these diseases are either monopartite or bipartite. Designated as new world begomoviruses, bipartite genomes contain DNA-A and DNA-B portions. Monopartite genomes, also known as old world (OW) begomoviruses, contain only one DNA-A fragment. However, bipartite Begomovirus DNA-A can sicken a person alone and be responsible for further propagation in OW. DNA-A in gammoviruses is dependent on DNA-B for proper operation. Notwithstanding parts, some DNA satellites (betasatellites and alpha satellites) are likewise present (Bridson and Stanley, 2006). Betasatellites are assistant of Begomoviruses and have ss-DNA (Bridson *et al.*, 2003). All capacities are done by CP. That CP has numerous capacities of inception and

pathogenicity (Bridson, 2003). Dissimilar to betasatellites, alphastellites can reproduce freely and have size ~1400 nt (Mansoor *et al.*, 1999). As per present day study, these are different in nature (Zulfiqar *et al.*, 2012). The genome size is 2.8 kb long. DNA-A segment have five to six replication sites in the two sides that encode 10kDa protein. Protein is associated with numerous capacities like replication, hushing and transmission and so forth (Bisaro, 2006). Because of appearance of new terminology for Begomoviruses, these infections could be arranged into a few animal groups including: Chilli leaf twist multan infection (ChiLCMV), Chilli leaf twist India infection (ChiLCINV), Chilli leaf twist Panipat (ChiLCPP), Chilli leaf twist Oman infection (ChiLCOV) and Pepper leaf twist Lahore infection (PepLCLV). Viruses like PepLCLV (JN135234, JN663864, and JX524173) have shown for disease significance. A few animal varieties spread in Pakistan as well as announced from different regions of the world, for example, Oman, Bangladesh, Sri Lanka and so forth. The genome of Begomoviruses ss-DNA gives an

[Citation: Atif, M., Ahmad, F., Manzoor, M.T., Gilani, K., Ali, Q., Sarwar, M., Anjum, S., Alam, M.W., Hussain, A., Rafaqat, N. (2022). Diversity, mutation and phylogenetic analysis of pepper leaf curl Lahore virus and pepper leaf curl Bangladesh virus infecting chilli crop Asia and China. *Biol. Clin. Sci. Res. J.*, 2022: 112. doi: <https://doi.org/10.54112/bcsrj.v2022i1.112>]



opportunity to decide their recombination and transformation rates. In this examination, transformative investigation of bean stew and pepper infections from various pieces of subcontinent have been provided. We have additionally created an information bank showing what succession was detached, when grouping was secluded, from where arrangement was confined.

**Methodology and Materials**

**Sequences Organization**

*PepLCLV* and *PepLCBV* complete sequences were gathered from National Center for Biotechnology Information. To compare the resemblance of different viruses (at the specie level), a BLAST is used (Muhire et al., 2014). Evolutionary dendrogram has been also given (Figure 1).

**Evolutionary Examination**

**Table 1:** Presence of various *PepLCV* in various locations.

Count	Pathogen	Registration Number	Location		Reference
I.	Pepper leaf curl	JN135234	India	India	GB file*
II.	Pepper leaf curl	JN663864	India	India	GB file
III.	Pepper leaf curl	JX524173	India	India	GB file
IV.	Pepper leaf curl	NC_016984	Pakistan	Pakistan	GB file
V.	Pepper leaf curl	AM691745	Pakistan	Pakistan	GB file
VI.	Pepper leaf curl	AM491589	Pakistan	Pakistan	GB file
VII.7	Pepper leaf curl	AM404179	Pakistan	Pakistan	GB file
VIII.	Pepper leaf curl	JN880419	Pakistan	India	GB file

\*GB file= Gene Bank file collected from NCBI.

**Phylogenetic analysis**

Eight complete genomes in all were examined. These genomes revealed the presence of four strains (SDT

Using the MUSCLE alignment approach, every feature length sequences were mapped in MEGA 6 software. The maximum likelihood approach is used to build the dendrogram (Kumar et al., 2016). Remarkably similar separates were turned into triangles via MEGA 6's tree editor. The *PepLCLV* have eight genomes (Table 1), *PepLCBV* have four genomes. Using the Sequence Demarcation Tool (SDT) application, bilateral similarity was calculated (Muhire et al., 2014).

**Results**

**Data bank analysis**

Eight sequences of *PepLCLV* were reported in subcontinent. Equal number of genomes was isolated in each country that is four. Four sequences of *PepLCBV* reported from Bangladesh (Table 1).

analysis). Only Pakistan and India were the countries that reported these (GB file).

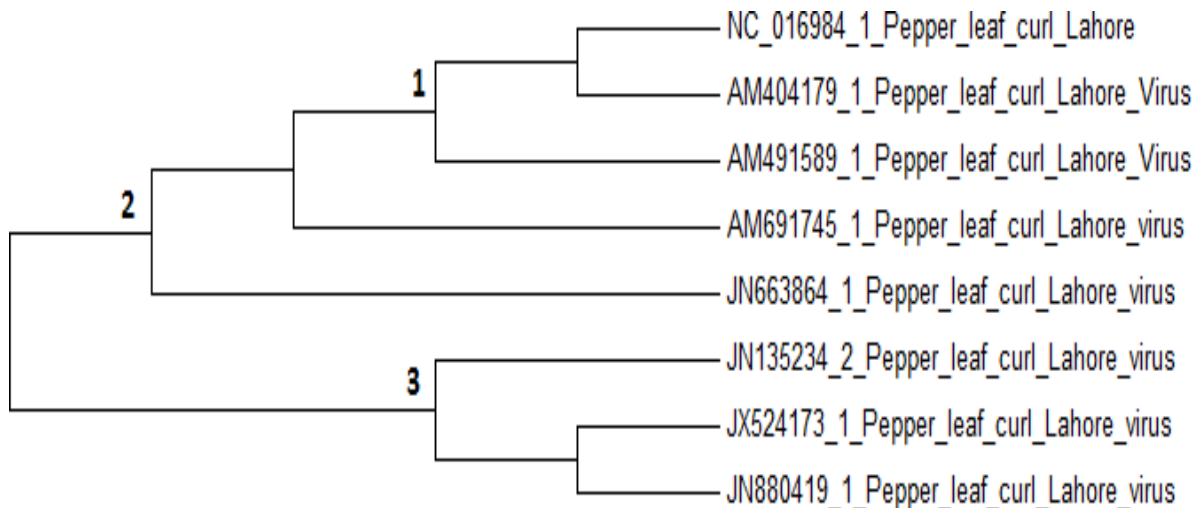


Figure 1. Dendrogram of *PepLCLV*

[Citation: Atif, M., Ahmad, F., Manzoor, M.T., Gilani, K., Ali, Q., Sarwar, M., Anjum, S., Alam, M.W., Hussain, A., Razaqat, N. (2022). Diversity, mutation and phylogenetic analysis of pepper leaf curl Lahore virus and pepper leaf curl Bangladesh virus infecting chilli crop Asia and China. *Biol. Clin. Sci. Res. J.*, 2022: 112. doi: <https://doi.org/10.54112/bcsrj.v2022i1.112>]

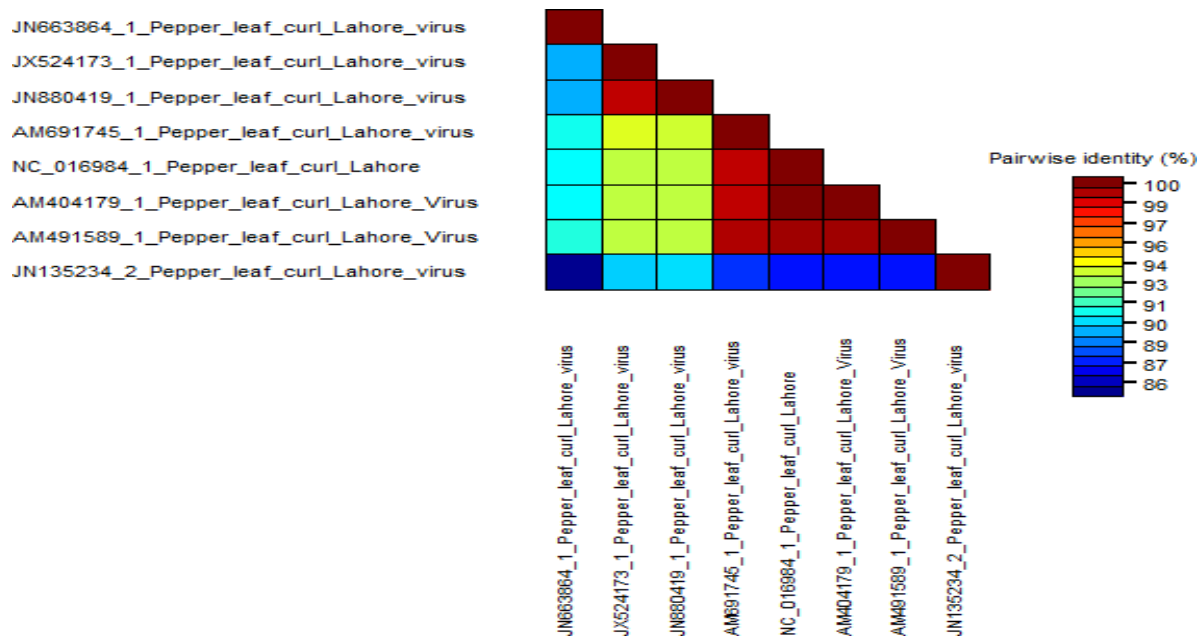


Figure 2. Grid (SDT Analysis) displaying evolution gap among *PepLCLV*. Grid values are maintained around 94 and 91 to identify species (New demarcation rule)

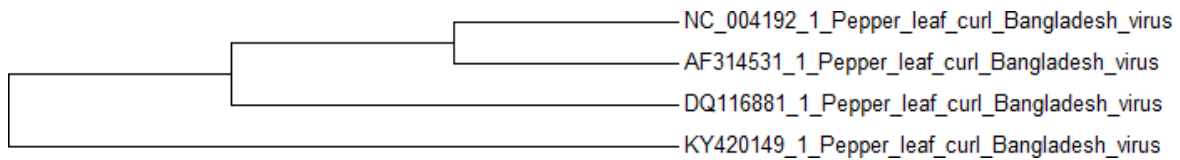


Figure 3. Dendrogram of *PePLCBV*

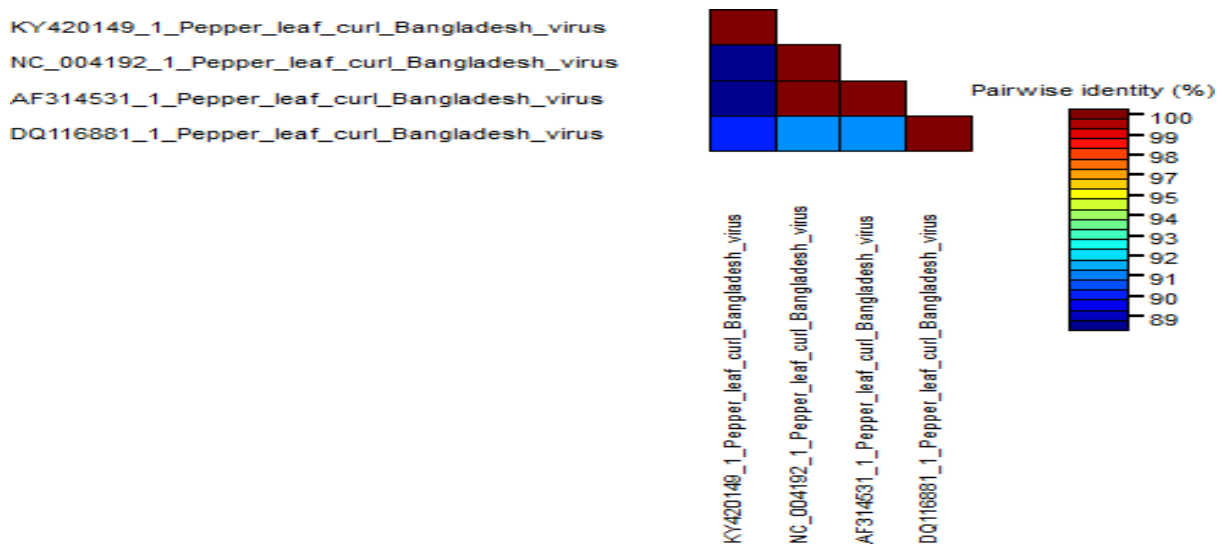


Figure 4. Grid (SDT Analysis) displaying evolution gap among *PePLCBV*. Grid values are maintained around 94 and 91 to identify species (New demarcation rule).

A dendrogram was used to evaluate a set of eight genomes. Other genomes were offspring of the one whose registration number was JN880419. The rest genomes were obtained in Pakistan, while those with the identification codes JN663864, JN135234, JN880419, and JX524173 were separated from India. These genomes contained four distinct strains. The first strain, which was identified as coming from Nagpur (India), had just one genome, whereas the second and third strains each have two, four, and three genomes, respectively (GB file). Lots of factors might be at play in this variation and dispersion. These factors might include the emergence of new insect vectors, the exchange of contaminated goods among those 2 nations, and more. Some of these diseases were also found in plants besides stew, including other plants. It demonstrates that this pathogen has a variety of hosts. Whenever a microbe has a lot of different hosts, there is always a good reason for it. Because of various vector species, infections could advance as indicated by their vectors and become contagious (Figure 1). Four arrangements were characterized in phylogenetic tree. Two groupings were disconnected from Bangladesh and two from Pakistan. This infection was just detailed from bean stew crop up until now. Three arrangements were segregated by Chinese analysts and just one grouping was presented by Pakistani scientists.

### Discussion

This examination has some remarkable focus when contrasted with past investigations. This investigation was showing changeability in *Pepper leaf twist infection* as species and strains. Our outcomes was showing huge variety. Our outcomes was showing change and recombination. That might be helpful at anticipating. That investigation might give development to next generation so that scientist may confine *Chili leaf twist infection* from different yields which was not influenced from this infection yet however are powerless against assault by this microorganism. More advancement could be possible on brinjal crop since it's anything but an equivalent group of stew crop. Hereditary variety is significant for huge populace sizes to exist (Biebricher and Eigen, 2006). The RNA infections transform quickly when contrasted with DNA infections (Holland, 1982). This was because of RNA infections give more destinations to hybridization (Worobey and Holmes, 1999; Eigen *et al.*, 1988). Nonetheless, if there was an occurrence of single-abandoned DNA infections, development is probably going to be happened all the more quickly when contrasted with RNA infections (Shackelton and Holmes, 2006; Duffy *et al.*, 2008). Infections have a place with *Geminiviridae* family showed tremendous hereditary variety. This was because of hybridization

(Grigoras *et al.*, 2010; Ge *et al.*, 2007). Infections using DNA polymerase were heritably more varied, according to studies (Duffy and Holmes, 2009). The secret to inherited variation is transformation, notwithstanding (Balol, 2010). Hybridization is responsible for the spread of plant diseases, especially Geminiviruses, according to studies (Lefurve and Moriones, 2015). Actually, their dependent reproductive portion was the source of this fusion (Jeske *et al.*, 2001). Due to the commitment of DNA fragments from different viruses, recombination occurred (Zhou *et al.*, 1997; Barrie *et al.*, 2001; Monci *et al.*, 2002). Hereditary changeability and variation were brought about by this recombination (Silva *et al.*, 2014). Our findings demonstrated that the exchange of sections across infections results in the creation of a new strain, adding to the enormous variety of viral population. Begomovirus's genetic diversity resulted from two variables: their increased value of genome substitution (Duffy and Holmes, 2008, 2009) and ongoing crossover, which promotes advancement (Padidam *et al.*, 1999; Pita *et al.*, 2001). Therefore, metamorphosis and mixing play a crucial role in the hereditary diversity and instability of begomoviruses. Bipartite Begomovirus *Tomato Serious Rugose Infection* (ToSRV) and *Macrotidium Yellow Spot Infection* were explored by Lima *et al.* in 2013. A few studies suggested that begomoviruses merged often for a very long time (Martin *et al.*, 2011). This resulted from the break focus's employment of a moving circular duplication instrument to replicate or improvement (Lefurve *et al.*, 2007a; Prasanna and Rai, 2007). Results from our expanded analysis revealed that some configurations (LN845958 and LN845957) were not associated to cotton. Because of the diversity of viral groups, these infections could only be transmitted to cotton as a host. Geminiviruses' coat proteins provide important information regarding mixing and hereditary diversity (Unsel *et al.*, 2001; Liu *et al.*, 1997). Transmission required the coat protein. A few duplication errors could also lead to changes at certain grouping points (Zhang *et al.*, 2001). Geminiviruses discovered across Brazil exhibited high hereditary variation and changeability through hybridization (Galvao *et al.*, 2003; Inoue-Nigata *et al.*, 2006; Ribeiro *et al.*, 2007). Through recombination, new strains that trigger pandemics were given at fresh yields. Weeds were the Begomoviruses' mixing station, according to studies (Castillo-Urquiza *et al.*, 2008; Roye *et al.*, 1999).

It was later established that weeds serve as a foundation for hybridization. Begomoviruses develop, diversify, and then infect Jamaica (Paprotka *et al.*, 2010b). Sections of the ssDNA of



Geminiviruses appear to cause genetic material to mutate, according to some data (Harkins *et al.*, 2009). In Brazil, numerous viruses, including the *BGMV*, exhibit little phenotypic mutations (Faria and Maxwell, 1999). However, when studies were carried out using the RCA method, this virus demonstrated increased heterogeneity within a species (Ramos-Sobrinho *et al.*, 2010). Multiple new and aggressive variants of *CLCuV* have entered the cotton crop, according to diversity analysis of *CLCuV*, and *CLCKoV* and *CLCuMuV* are to blame for this (Ahmed, 2010; Ahmad, 2011; Briddon, 2003; Di Carli, 2012; Duffy, 2008; Ahmad, 2011; Bosco, 2004).

A begomovirus known as *PeLCV* infects 15 distinct host, including *Pedilanthus*. These hosts are primarily from Pakistan. According to certain investigations, it is evolved and originated in Pakistan (Moriones and Castillo, 2000; Zaidi *et al.*, 2016). *Cucumber mosaic virus* (CMV), for example, has been found to infect a number of host plants (Palukaitis and Garca-Arenal, 2003). One of the finest viruses from the Geminivirus family to infect a variety of plants is the *pepper leaf curl virus* (PeLCV) (Shakir *et al.*, 2018). The development or appearance in leaf curl infection are caused by *CLCuKoV* (Saleem *et al.*, 2016). Research has demonstrated that begomovirus Rep genes are more translational and diverse than Cp genes (Lima *et al.*, 2017; Zhou *et al.*, 1997; Fondong *et al.*, 2000; Patil and Fauquet, 2009). *EACMV* and *ACMV* are joined in South Africa and Angola, resulting in a novel variety of the begomovirus in both nations (Kumar *et al.*, 2009; Bisimwa *et al.*, 2012; Mabora *et al.*, 2008; Cossa, 2010). *Begomoviruses* are major cause of leaf curl diseases in India in chilli crop (Kumar *et al.*, 2015). Combine attack of *ChiLCV* and *PepLCBV* has also been seen (Kumar *et al.*, 2015). This is also found at six different places in India (Nagpur, Palampur, Ghazipur, Salem, New Delhi and Chapra) (Kumar *et al.*, 2015).

### Conclusion

*Pepper leaf curl virus* was a highly virulent virus. It was responsible for crop losses across subcontinent. Evolution played important role in its diversity. It had a strong ability to recombine with other viruses to evolve into a new strain. This study indicated that the spread of this virus into different areas of subcontinents was excellent.

### Acknowledgement

Author acknowledges the facilities given by department of Plant Pathology, faculty of agricultural sciences to complete this work.

### Conflict of interest

The author declared absence of conflict of interest.

### References

- Ahmed. (2010). Phylogenetic analysis of *Bemisia tabaci* (Hemiptera: Aleyrodidae) populations from cotton plants in Pakistan, China, and Egypt. *Journal of Pest Science*, **14**, 135-141.
- Ahmad. (2011). Characterization of *Sun hemp begomo virus* and its geographical origin based on in silico structural and functional analysis of recombinant coat protein. *African Journal of Biotechnology*, **10**, 260-270.
- Balol, G.B. (2010). Sources of genetic variation in plant virus populations. *Journal of Pure Applied Microbiology* **4**, 803-808.
- Berrie, L.C., Rybicki, E.P., Rey, M.E.C. (2001). Complete nucleotide sequence and host range of South African cassava mosaic virus: further evidence for recombination amongst *Begomoviruses*. *Journal of General Virology*, **82**, 53-58.
- Biebricher, C.K., & Eigen, M. (2006). 'What is a Quasispecies?'. *Current Topics in Microbiology and Immunology*, **299**, 1-31.
- Bisaro. (2006). Silencing suppression by *Geminivirus* proteins. *Virology*, **16**, 158-168.
- Bisimwa, E., Walangululu J., & Bragard, C. (2012). Occurrence and Distribution of *cassava mosaic begomovirus* related to agro-ecosystems in the Sud-kivu Province, Democratic Republic of Congo. *Asian Journal of Plant Pathology*, **6**, 1-12. doi: 10.3923/ajppaj.2012.1.12.
- Bosco. (2004). *TYLCSV* DNA, but not infectivity can be transovarially inherited by the progeny of the whitefly vector *Bemisia tabaci* (Gennadius). *Virology*, **18**, 276-283.
- Briddon. (2003). Diversity of DNA  $\alpha$  : a satellite molecule associated with some monopartite *begomoviruses*. *Virology*, 106-121.
- Briddon, R.W., Bull, S.E., Amin, I., Idris, A.M., Mansoor, S., Bedford, I.D., & Markham, P.G. (2003). Diversity of DNA  $\beta$ , a satellite molecule associated with some monopartite *Begomoviruses*. *Virology*, **312**, 106-121.
- Briddon, R.W., & Stanley, J. (2006). Subviral agents associated with plant single-stranded DNA viruses. *Virology*, **344**, 198-210.
- Castillo-Urquiza, G.P., Beserra, J.E.A., Bruckner, F.P., Lima, A.T., Varsani, A., Alfenas-Zerbini, P., & Zerbini, F.M. (2008). Six novel begomoviruses infecting tomato and associated weeds in Southeastern Brazil. *Archives of virology*, **153**(10), 1985-1989.
- Duffy, S., & Holmes, E.C. (2008). Phylogenetic evidence for rapid rates of molecular evolution in the single-stranded DNA begomovirus *Tomato yellow leaf curl virus*. *Journal of General Virology*, **82**, 957-965.
- Duffy, S., & Holmes, E.C. (2009). Validation of high rates of nucleotide substitution in *geminiviruses*:

[Citation: Atif, M., Ahmad, F., Manzoor, M.T., Gilani, K., Ali, Q., Sarwar, M., Anjum, S., Alam, M.W., Hussain, A., Rafaqat, N. (2022). Diversity, mutation and phylogenetic analysis of pepper leaf curl Lahore virus and pepper leaf curl Bangladesh virus infecting chilli crop Asia and China. *Biol. Clin. Sci. Res. J.*, **2022**: 112. doi: <https://doi.org/10.54112/bcsrj.v2022i1.112>]

- phylogenetic evidence from east african *cassava mosaic viruses*. *Journal of General Virology*, **90**, 1539-1547.
- Duffy, S., Shackelton, L.A., & Holmes, E.C. (2008). Rates of evolutionary change in viruses: patterns and determinants. *Nature Reviews Genetics*, **9**, 267-276.
- Eigen, M., Winkler-Oswatitsch, R., Dress, A. (1988). Statistical geometry in sequence space: a method of quantitative comparative sequence analysis. *Science USA*, **85**, 5913-5917.
- Faria, J.C., & Maxwell, D.P. 1999. Variability in geminivirus isolates associated with *Phaseolus* spp. in Brazil. *Phytopathology*, **89**, 262-268.
- Fondong, V.N., Pita, J.S., Rey, M.E.C., de Kochko, A., Beachy, R.N., & Fauquet, C.M. (2000). Evidence of synergism between *African cassava mosaic virus* and a new double-recombinant geminivirus infecting cassava in Cameroon. *Journal General Virology*, **81**, 287-297.
- Galvao, R.M., Mariano, A.C., & Luz, D. (2003). A naturally occurring recombinant DNA-A of a typical bipartite begomovirus does not require the cognate DNA-B to infect *Nicotiana benthamiana* systemically. *Journal General Virology*, **84**, 715-726.
- Ge, L., Zhang, J., Zhou, X., & Li, H. (2007). Genetic structure and population variability of *tomato yellow leaf curl China virus*. *Journal of virology*, **81**(11), 5902-5907.
- Grigoras, I., Timchenko, T., Grande-Pérez, A., Katul, L., Vetten, H.J., & Gronenborn, B. (2010). High variability and rapid evolution of a *nanovirus*. *Journal of virology*, **84**(18), 9105-9117.
- Harkins, G.W., Delpont, W., & Duffy, S. (2009). Experimental evidence indicating that *mastreviruses* probably did not co-diverge with their hosts. *Virology Journal*, **6**, 104.
- Holland, J., Spindler, K., Horodyski, F., Grabau, E., Nichol, S., & Vande Pol, S. (1982). Rapid evolution of RNA genomes. *Science*, **215**(4540), 1577-1585.
- Inoue-Nagata, A.K., Martin, D.P., Boiteux, L.S., Giordano, L.D., Bezerra, I.C., & De Avila, A.C. (2006). New species emergence via recombination among isolates of the *Brazilian tomato infecting Begomovirus* complex. *Brazilian Journal of Agricultural Research*, **41**, 1329-1332.
- Jeske, H., Lutgemeier, M., & Preiss, W. (2001). DNA Forms Indicate Rolling Circle and Recombination-Dependent Replication of *Abutilon Mosaic Virus*, *EMBO Journal*, **20**: 6158-6167.
- Kumarvinoth, Tribhuvan, Y.V., & SaumikBasu. (2015). Complexity of *Begomovirus* and betasatellite populations associated with chili leaf curl disease in India. *Journal of General Virology*, **96**, 3143-3158.
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology Evolution*, **33**, 1870-1874. doi: 10.1093/molbev/msw054.
- Lefevre, P., Lett, J.M., Reynaud, B., Martin, D.P. (2007a). Avoidance of protein fold disruption in natural virus recombinants. *PLoS Pathology*, **3**:e181.
- Lefevre., & Moriones. (2015). Recombination as a motor of host switches and virus emergence: *Geminiviruses* as case studies. *Current Opinion in Virology*, **10**, 14-19.
- Lima, A. T., Sobrinho, R. R., Gonzalez-Aguilera, J., Rocha, C. S., Silva, S. J., Xavier, C. A., & Zerbini, F. M. (2013). Synonymous site variation due to recombination explains higher genetic variability in begomovirus populations infecting non-cultivated hosts. *Journal of General Virology*, **94**(2), 418-431.
- Lima, A., Silva, J. C., Silva, F. N., Castillo-Urquiza, G. P., Silva, F. F., Seah, Y. M., & Zerbini, F. M. (2017). The diversification of begomovirus populations is predominantly driven by mutational dynamics. *Virus evolution*, **3**(1).
- Liu, H., Boulton, M. I., & Davies, J. W. (1997). *Maize streak virus* coat protein binds single- and double-stranded DNA in vitro. *Journal of general virology*, **78**(6), 1265-1270.
- Mansoor, S., Khan, S.H., Bashir, A., Saeed, M., Zafar, Y., Malik, K.A., & Markham, P.G. (1999). Identification of a novel circular single-stranded DNA associated with cotton leaf curl disease in Pakistan. *Virology*, **259**, 190-199.
- Martin, D.P., Biagini, P., Lefevre, P., Golden, M., Roumagnac, P., & Varsani, A. (2011). Recombination in eukaryotic single stranded DNA viruses. *Viruses*, **3**, 1699-1738.
- Monci, F., Sánchez-Campos, S., Navas-Castillo, J., & Moriones, E. (2002). A natural recombinant between the geminiviruses *Tomato yellow leaf curl Sardinia virus* and *Tomato yellow leaf curl virus* exhibits a novel pathogenic phenotype and is becoming prevalent in Spanish populations. *Virology*, **303**(2), 317-326.
- Moriones, E., & Navas-Castillo, J. (2000). *Tomato yellow leaf curl virus*, an emerging virus complex causing epidemics worldwide. *Virus Research*, **71**, 123-134.
- Muhire, B. M., Varsani, A., & Martin, D. P. (2014). SDT: a virus classification tool based on pairwise sequence alignment and identity calculation. *PloS one*, **9**(9), e108277.

[Citation: Atif, M., Ahmad, F., Manzoor, M.T., Gilani, K., Ali, Q., Sarwar, M., Anjum, S., Alam, M.W., Hussain, A., Rafaqat, N. (2022). Diversity, mutation and phylogenetic analysis of pepper leaf curl Lahore virus and pepper leaf curl Bangladesh virus infecting chilli crop Asia and China. *Biol. Clin. Sci. Res. J.*, **2022**: 112. doi: <https://doi.org/10.54112/bcsrj.v2022i1.112>]

- Padidam, M., Sawyer, S., & Fauquet, C. M. (1999). Possible emergence of new geminiviruses by frequent recombination. *Virology*, **265**(2), 218-225.
- Palukaitis., & García-Arenal F. (2003). *Cucumoviruses. Advances in Virus Research*, **62**, 241-323.
- Paprotka, T., Metzler, V., & Jeske, H. (2010b). The first DNA 1-like alpha satellites in association with New World begomoviruses in natural infections. *Virology*, **404**, 148-157.
- Pita, J. S., Fondong, V. N., Sangare, A., Otim-Nape, G. W., Ogwal, S., & Fauquet, C. M. (2001). Recombination, pseudorecombination and synergism of geminiviruses are determinant keys to the epidemic of severe cassava mosaic disease in Uganda. *Journal of General Virology*, **82**(3), 655-665.
- Patil, B.L., & Fauquet, C.M. (2009). *Cassava mosaic geminiviruses: Actual knowledge and perspectives. Molecular Plant Pathology*, **10**, 685-701.
- Prasanna, H.C., & Rai, M. (2007). 'Detection and Frequency of Recombination in Tomato-Infecting Begomoviruses of South and Southeast Asia'. *Virology Journal*, **4**, 111.
- Ramos-Sobrinho, R., Silva, S.J.C., & Silva, T.A.L. (2010). Genetic structure of a population of the begomovirus Bean golden mosaic virus (BGMV) that infects lima bean (*Phaseolus lunatus L.*) in the state of Alagoas, Brazil. In: Program and Abstracts, 6<sup>th</sup> International Geminivirus Symposium and 4th International ssDNA Comparative *Virology Workshop.*, 2010, Guanajuato, Mexico.
- Ribeiro, S.G., Martin, D.P., Lacorte, C., Simoes, I.C., Orlandini, D.R.S., & Inoue-Nagata, A.K. (2007). Molecular and biological characterization of *Tomato chlorotic mottle virus* suggests that recombination underlies the evolution and diversity of Brazilian tomato begomoviruses. *Phytopathology*, **97**, 702-711.
- Roye, M.E., Spence, J., McLaughlin, W.A., & Maxwell, D.P. (1999). The common weed *Macroptilium lathyroides* is not a source of crop-infecting geminiviruses from Jamaica. *Tropical Agriculture*, **76**, 256-262.
- Saleem, H., Nahid, N., Shakir, S., Ijaz, S., Murtaza, G., Khan, A. A., & Nawaz-ul-Rehman, M. S. (2016). Diversity, mutation and recombination analysis of cotton leaf curl geminiviruses. *PLoS One*, **11**(3), e0151161.
- Shackelton, & Holmes, E.C. (2006). Phylogenetic Evidence for the Rapid Evolution of Human B19 Erythrovirus. *Journal of Virology*, **80**, 3666-3669.
- Shakir, S., Nawaz-ul-Rehman, M.S., & Mubin, M. (2018). Characterization, phylogeny and recombination analysis of *Pedilanthus leaf curl virus*-Petunia isolate and its associated betasatellite. *Virology Journal*, **15**, 134 <https://doi.org/10.1186/s12985-018-1047-y>.
- Silva, F.N., Lima, A.T., Rocha, C.S., Castillo-Urquiza, G.P., Alves-Júnior, M., & Zerbini, F.M. (2014). Recombination and pseudorecombination driving the evolution of the begomoviruses *Tomato severe rugose virus* (ToSRV) and *Tomato rugose mosaic virus* (ToRMV): two recombinant DNA-A components sharing the same DNA-B. *Virology journal*, **11**(1), 1-11.
- Unsel, S., Höhnle, M., Ringel, M., & Frischmuth, T. (2001). Subcellular targeting of the coat protein of *African cassava mosaic geminivirus*. *Virology*, **286**(2), 373-383.
- Worobey, M., & Holmes, E.C. (1999). Evolutionary Aspects of Recombination in RNA Viruses. *Journal of General Virology*, **80**, 2535-2545.
- Zaidi, S.S., Martin, D.P., Amin, I., Farooq, M., & Mansoor, S., (2016). *Tomato leaf curl New Delhi virus*; a widespread bipartite begomovirus in the territory of monopartite begomoviruses. *Molecular Plant Pathology*, 2016.
- Zhang, Wei., Olson, N.H., Baker, T.S., Faulkner, L., Agbandje-McKenna, M., Boulton, M.I., Davies, J.W., & McKenna, R. (2001). Structure of the *Maize streak virus* geminate particle. *Virology*, **279**, 471-477.
- Zhou, X., Liu, Y., Calvert, L., Munoz, C., Otim-Nape, G.W., Robinson, D.J., & Harrison, B.D. (1997). Evidence that DNA-A of a geminivirus associated with severe cassava mosaic disease in Uganda has arisen by interspecific recombination. *Journal General Virology*, **78**(8), 2101-2111.
- Zulfiqar, A., Zhang, J., Cui, X., Qian, Y., Zhou, X., & Xie, Y. (2012). A new *Begomovirus* associated with alpha- and betasatellite molecules isolated from *Vernonia cinerea* in China. *Archives of Virology*, **25**, 189-191.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this

article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022